

An *Escherichia coli* gene showing a potential ancestral relationship to the genes for the mitochondrial import site proteins ISP42 and MOM38

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An ORF (*OrfT*) of 1911 base pairs, upstream of the *hip* operon in *Escherichia coli* at map position 33.82 has been identified. The protein encoded by this sequence is predicted to have a molecular mass of 68 249 Da and the carboxyterminal 276 residues shows 26.8% and 25.4% identity with the import site proteins ISP42 and MOM38 from the mitochondrial outer membrane of *Saccharomyces cerevisiae* and *Neurospora crassa*, respectively. These mitochondrial membrane proteins have been shown to be essential components of the protein translocation apparatus in yeast. These similarities raise the possibility that *OrfT* might represent the bacterial gene from which these eukaryotic genes evolved.

It is now widely accepted that, in the course of evolution of the eukaryotic cell, mitochondria have evolved from endosymbiotic bacteria [1]. The supposed evolution of the mitochondria requires that a system for protein import be established in the outer membrane of the ancestral endosymbiont, since most of the proteins in mitochondria are now nuclear-encoded and synthesized in the cytosol. The mitochondrial protein import machinery is localized in the mitochondrial outer membranes and consists of a number of proteins which include receptor-like components and import site proteins, some of which have now been identified in the fungi, *Saccharomyces cerevisiae* and *Neurospora crassa* [2]. One of the import site proteins (ISP42 in yeast [3]; MOM38 in *Neurospora* [4]) has been shown to be essential for cell viability. A number of genes involved in protein translocation in *E. coli* have also been identified (reviewed in Ref. 5). Surprisingly, to date there is little evidence for homology between components of the bacterial and mitochondrial translocation machinery [2].

The yeast and *Neurospora* outer membrane proteins ISP42 and MOM38 show an overall sequence homology of 53.2%, but in some regions almost 100% identity is observed (Fig. 3). Based on the assumption that

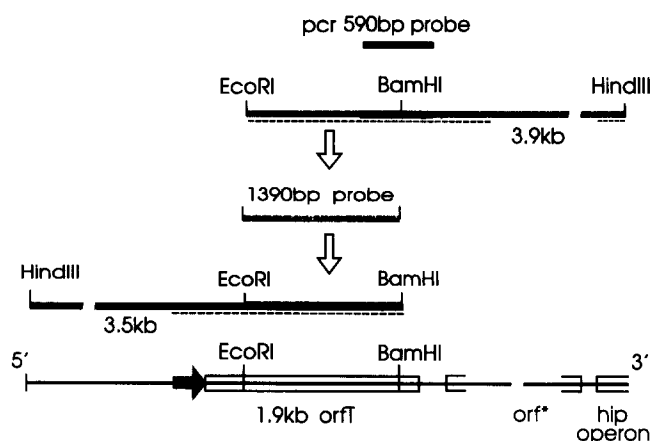


Fig. 1. Relationship of *OrfT* to the *hip* locus of *E. coli*. The overlapping 3.9 kb *EcoRI*-*HindIII* and 3.5 kb *HindIII*-*BamHI* fragments contained the 1.9 kb *OrfT*. The *hip* operon is between 3.8 and 3.9 kb downstream of the initiating ATG in *OrfT*, at map position 33.90. A second open reading frame (*orf**) is located between *OrfT* and *hip*, but only the 5' and 3' ends of this *orf* has been sequenced. The promoter elements identified for *OrfT* are shown by the solid arrow. The portions of the *E. coli* DNA sequenced is shown by (---). The 590 bp probe, generated by PCR, used to isolate the 3.9 kb fragment and the 1396 bp *EcoRI*-*BamHI* fragment used to obtain upstream sequences for *OrfT* are also shown.

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these highly conserved regions are essential for the structure and function of these proteins, oligonucleotide primers were designed corresponding to two such regions (ISP42 residues 126–134 and 300–309) and these were used in a PCR, with *E. coli* chromosomal DNA template, to produce a 590 bp probe. This probe was subsequently used to screen an *E. coli* genomic library and a fragment of approx. 3900 bp was isolated, bounded by unique *Eco*RI and *Hind*III sites. A 1390 bp *Eco*RI-*Bam*HI subfragment from the 5' end of the 3.9 kb clone was used to obtain an overlapping fragment of approx. 3.5 kb with *Hind*III-*Bam*HI ends (Fig. 1). Nucleotide sequence was obtained for the overlapping fragments from approx. 600 bp upstream of the *Eco*RI site to approx. 2.2 kb downstream of the *Eco*RI site (Fig. 1). The sequence contained an ORF of 1911 base pairs (*OrfT*) (Fig. 2) and the start of a second ORF approx. 44 bp downstream from *OrfT*. Using DNase to prepare deletion fragments the nucleotide sequence at the 3' end of the 3.9 kb clone was determined and revealed that the fragment extends into the

hip locus [6] which has been mapped to position 33.90 on the *E. coli* chromosome [7]. Since this locus is between 3.8 and 3.9 kb downstream of *OrfT*, we propose that the map position of *OrfT* is 33.82. The initiation codon ATG (at position 1, Fig. 2) is preceded by a ribosome binding site (AGG) and promoter elements at positions –35 (TTGGCG) and –10 (TTTAAT) with respect to the transcription start site which show the basic characteristics of *E. coli* promoters [8]. There are no obvious promoter elements between *OrfT* and the second ORF, raising the possibility that these two genes are polycistronic.

A search of the EMBL/GenBank Data Libraries revealed that part of the *OrfT* sequence (from residue 1740 to 2130) shows 98.0% identity with a fragment of DNA from landmark loops of lampbrush chromosomes from the amphibian newt, *Pleurodeles waltlii* [9]. Since the *OrfT* sequence is contiguous with that of the *E. coli hip* operon, it appears likely that the newt sequence is in fact an *E. coli* sequence and therefore a cloning artefact. The 1911 bp *OrfT* sequence encodes

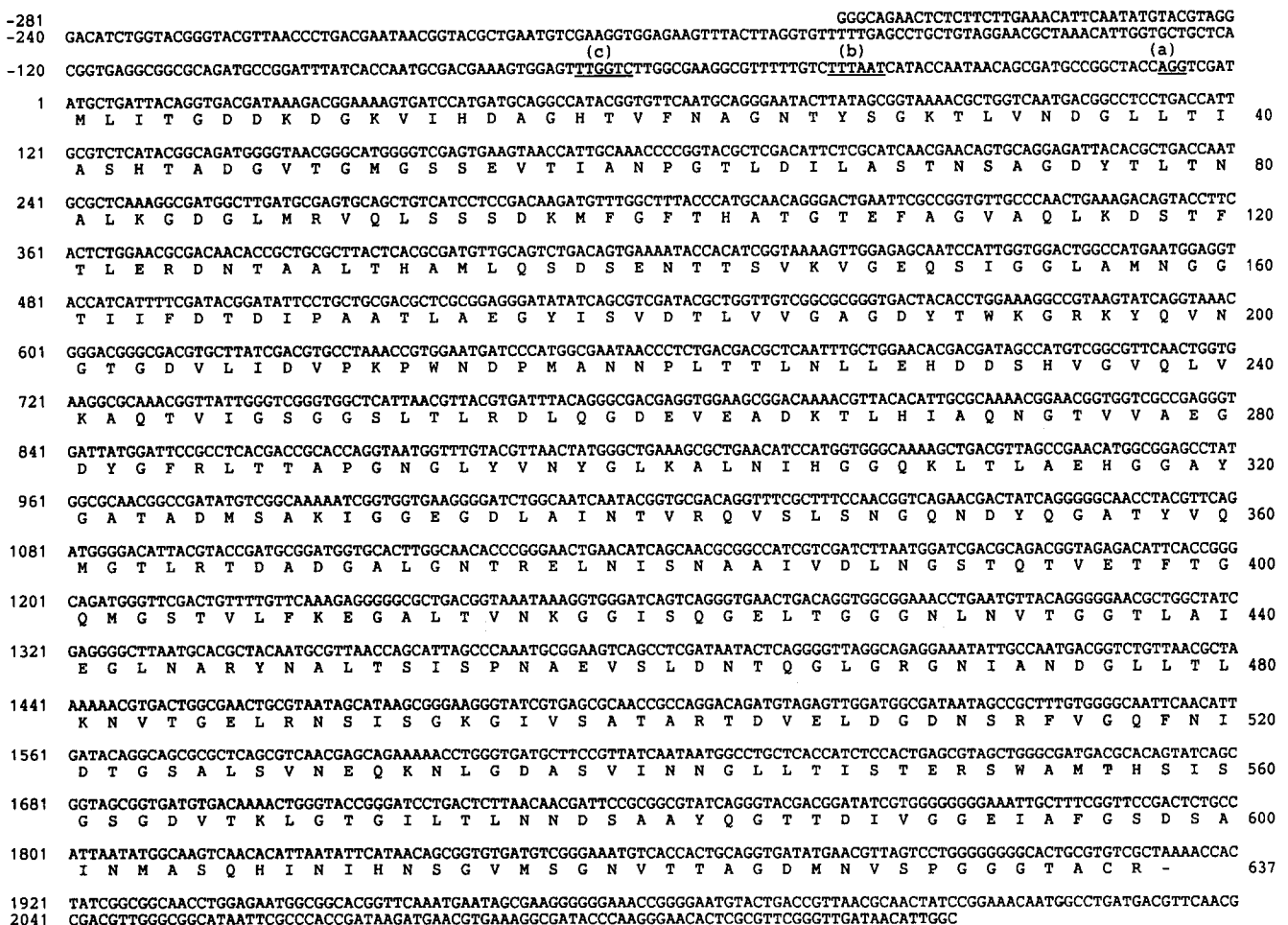


Fig. 2. Nucleotide sequence and deduced amino acid composition of *OrfT*. The translated sequence is given in the single letter code. The numbering of nucleotides is shown on the left hand side and amino acid residues on the right hand side. The initiation codon at position 1 is preceded by a putative ribosome binding site (a) and promoter elements at –10 (b) and –35 (c). The potential transcription start site is approx. 10 bases down-stream of (b).

ISP42	IPTIPALSPITAKQSKGNFSSNPISSEFVVDYKQLHSHR	53
MOM38	MASFSTESPLAMLRDNI-YSSLS-----DAFNAFQERR	33
OrfT	MGTERTDADGALGNTRRL-NISNYAA--IVDLNGSTQTV-	394
ISP42	QSLELVNPGTVENLNKEVSRDVFLSQYFFTGRLADLNKAF	93
MOM38	KQFGLSNPGTIETIAREVORDTLTNYMFSGLRADVTKAF	73
OrfT	-ETFTGQMGSTV-LFKEGALTVNKGGISQGEETGGGNLV	432
ISP42	SMNFAFQTSTHSISGSQALPKYAFSALFANDNLFAQGNID	133
MOM38	SLAPLFQVSHQFAMGER-LNPIYAFALYGTNQIFAQGNLD	112
OrfT	TGGTLAIEGLNARYNALTSISPNAEVSLEDTQGLGRGNIA	472
ISP42	NDLSVSGRLNYGWDKKNISKVNLQISDGGPTMCQLEQDYQ	173
MOM38	NEGALSTRFNRYRWGDRITITKQFSIGGGDMA-QFEHEHL	151
OrfT	NDGLLTLK-NVTGELRNSISGKGIVSATARTDVELDGD-N	510
ISP42	ASDFSVMVKTLPNPFSEKGEFTVGAVAS-FLQS-VTFOLA	211
MOM38	GDDFSASLKAINPSFLD-GGLTGIFVGD-YLQA-VTERLG	188
OrfT	-SREVGQFNIDTGSALSVDNQKNGDASVINNGLLTISTE	549
ISP42	-GLETLYSRTDGSAPGDAGVSYLTRYVSKQDWIFSGQLQ	250
MOM38	LGLQAVWQRCGLTQGPDTAISYFARY--KAGDWVASAQLQ	226
OrfT	RSWAMTHSISGSDVTKLGTGILTLLNDSAA-YQGTDTIV	588
ISP42	-ANGALIAS-LWRKVAQNVEAGIE--TTLQAGMVPITDPL	286
MOM38	-AQGALNTS-FWKKLTDVQAGVD--HTLSVAP---SQSM	259
OrfT	GGETAFGSDSAINMASQIHINHNHNSGVMSQNVTT--AGD-N	626
ISP42	MGTPIGIQPTV	297
MOM38	MG-----GLTK	265
OrfT	NVSPGGGTACR	637

Fig. 3. Amino acid similarities between *E. coli* ORFT, yeast ISP42 and *Neurospora* MOM38. Sequences starting with residue 361 of OrfT, 13 of ISP42 and 1 of MOM38 were compared. Identical residues and conservative replacements defined as T/S, D/E, K/R/H, I/L/V, G/A and Q/N are shaded grey.

a protein of 637 amino acids (Fig. 2) with an estimated molecular mass of 68 249 Da. The protein is very acidic, having a predicted pI of 4.26. Comparison of the protein sequence encoded by *OrfT* (ORFT) with ISP42 from *S. cerevisiae* and MOM38 from *N. crassa* shows that the carboxyterminal half of the molecule (residues 361 to 637) is 26.8% and 25.4% homologous, respectively (Fig. 3). This compares with a similarity of 53.2%, between ISP42 and MOM38. As is the case

with the integral membrane protein ISP42, ORFT also has no obvious α -helical transmembrane domains.

ORFT represents the first example of a bacterial protein with a similarity to the membrane components of the mitochondrial translocation machinery. In the absence of functional data, we cannot assume that the protein encoded by *OrfT* functions in bacterial protein translocation, but the similarity between ORFT and ISP42 and MOM38 suggests a potential relationship between these genes. We believe that the identification of *OrfT* provides further support for the hypothesis that genes from an endosymbiont ancestor were translocated by the host cell to create a system for protein import into mitochondria.

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